ml extract as explained above. MN proteins were found in endometrial (lanes D and M), ovarian (lanes E and N) and in uterine cervical (lane O) carcinomas. In those extracts MN-related proteins were found in three bands having molecular weights between about 48 kd and about 58 kd. Another MN-related protein was present in the tissue extract from a mammary papilloma; that protein was seen as a single band at about 48 kd (lane J).

Clearly negative were the extracts from full-term placenta (lane B), normal mammary gland (lane K), hyperplastic endometrium (lane L), normal ovaries (lane H), and from uterine myoma (lane I). Only extremely slightly MN-related bands were seen in extracts from trophoblasts (lanes F and G) and from melanoma (lane P).

The observations that antigen related to p54/58N was expressed in clinical specimens of several types of human carcinomas but not in general in normal tissues of the corresponding organs (exceptions delineated in Example 13) further strengthened the association of MN antigen with tumorigenesis. However, it should be noted that for human tumors, a normal tissue is never really an adequate control in that tumors are believed not to arise from mature, differentiated cells, but rather from some stem cells, capable of division and of differentiation. In body organs, such cells may be quite rare.

Example 7

MN Antigen in Animal Cell Lines

Since the MN gene is present in the chromosomal DNA of all vertebrate species that were tested, MN-related antigen was searched for also in cell lines derived from normal tissues and from tumors of several animal species. MN-related protein was found in two rat cell lines: one of them was the XC cell line derived from rat rhabdomyosarcoma induced with Rous sarcoma virus; the other was the Rat2-Tk$^-$ cell line. In extracts from both of those rat cell lines, a single protein band was found on the blots : its molecular weight on blots produced from a reducing gel and from a non-reducing gel was respectively 53.5 kd and 153 kd. Figure 10 shows the results with Rat2-Tk$^-$ cell extracts (lane B), compared with extracts from MX-infected HeLa (lane A); the concentration of MN antigen in those two cell lines is very similar. The extracts were analysed directly (40 μl per lane).

MN-related protein from XC cells showed the same pattern as for Rat2-Tk$^-$ cells both under reducing and non-reducing conditions, except that its concentration was about 30x lower. The finding of a MN-related protein--p53.5N--in two rat cell lines (Figures 10 and 12) provides the basis for a model system.

None of the other animal cell lines tested contained detectable amounts of MN antigen, even when the

highly sensitive immunoblot technique in which the MN antigens are concentrated was used. The MN-negative cells were: Vero cells (African green monkey); mouse L cells; mouse NIH-3T3 cells normal, infected with Moloney leukemia virus, or transformed with Harvey sarcoma virus; GR cells (mouse mammary tumor cells induced with MTV), and NMG cells (normal mouse mammary gland).

Example 8

Radioimmunoassays in Liquid Phase Using Recombinant MN Protein for MN-Specific Antibodies and for MN Antigen

The genetically engineered MN protein fused with glutathione S-transferase--GEX-3X-MN--prepared and purified as described above was labeled with ^{125}I by the chloramine T method [Hunter (1978)]. The purified protein enabled the development of a quantitative RIA for MN-specific antibodies as well as for MN antigens. All dilutions of antibodies and of antigens were prepared in RIPA buffer (1% TRITON X-100 and 0.1% sodium deoxycholate in PBS--phosphate buffered saline, pH 7.2), to which was added 1% of fetal calf serum (FCS). Tissue and cell extracts were prepared in RIPA buffer containing 1 mM phenylmethylsulfonylfluoride and 200 trypsin inhibiting units of Trasylol (aprotinin) per ml, with no FCS. ^{125}I-labeled GEX-3X-MN protein (2.27 $\mu Ci/\mu g$ of TCA-precipitable protein) was before use diluted with RIPA + 1% FCS, and non-specifically binding radioactivity was

adsorbed with a suspension of fixed protein A-Staphylococcus aureus cells (SAC).

In an RIA for MN-specific antibodies, MAb-containing ascites fluids or test sera were mixed with ^{125}I-labeled protein and allowed to react in a total volume of 1 ml for 2 hours at room temperature. Subsequently, 50 µl of a 10% suspension of SAC [Kessler, supra] was added and the mixture was incubated for 30 minutes. Finally, the SAC was pelleted, 3x washed with RIPA, and the bound radioactivity was determined on a gamma counter.

Titration of antibodies to MN antigen is shown in Figure 11. Ascitic fluid from a mouse carrying M75 hybridoma cells (A) is shown to have a 50% end-point at dilution 1:1.4 x 10^{-6}. At the same time, ascitic fluids with MAbs specific for MX protein (M16 and M67) showed no precipitation of ^{125}I-labeled GEX-3X-MN even at dilution 1:200 (result not shown). Normal rabbit serum (C) did not significantly precipitate the MN antigen; rabbit anti-MaTu serum (B), obtained after immunization with live MX-infected HeLa cells, precipitated 7% of radioactive MN protein, when diluted 1:200. The rabbit anti-MaTu serum is shown by immunoblot in Example 4 (above) to precipitate both MX and MN proteins.

Only one out of 180 human sera tested (90 control and 90 sera of patients with breast, ovarian or uterine cervical cancer) showed a significant precipitation of the

radioactively labeled MN recombinant protein. That serum--L8--(D) was retested on immunoblot (as in Example 4), but it did not precipitate any p54/58N from MX-infected HeLa cells. Also, six other human sera, including KH (E), were negative on immunoblot. Thus, the only positive human serum in the RIA, L8, was reactive only with the genetically engineered product, but not with native p54/58N expressed by HeLa cells.

In an RIA for MN antigen, the dilution of MAb M75, which in the previous test precipitated 50% of maximum precipitable radioactivity (= dilution 1:1.4 x 10^{-6}) was mixed with dilutions of cell extracts and allowed to react for 2 hours. Then, ^{125}I-labeled GEX-3X-MN (25 x 10^3 cpm/tube) was added for another 2 hours. Finally, the radioactivity bound to MAb M75 was precipitated with SAC and washed as above. One hundred percent precipitation (= 0 inhibition) was considered the maximum radioactivity bound by the dilution of MAb used. The concentration of the MN antigen in the tested cell extracts was calculated from an inhibition curve obtained with "cold" GEX-3X-MN, used as the standard (A in Figure 12).

The reaction of radioactively labeled GEX-3X-MN protein with MAb M75 enabled us to quantitate MN antigen directly in cell extracts. Figure 12 shows that 3 ng of "cold" GEX-3X-MN (A) caused a 50% inhibition of precipitation of "hot" GEX-3X-MN; an equivalent amount of MN

antigen is present in 3 x 10^3 ng of proteins extracted from MaTu-infected HeLa (B) or from Rat2-Tk$^-$ cells (C). Concentrations of MN protein in cell extracts, determined by this RIA, are presented in Table 2 below. It must be understood that the calculated values are not absolute, since MN antigens in cell extracts are of somewhat different sizes, and also since the genetically engineered MN protein is a product containing molecules of varying size.

TABLE 2

Concentration of MN Protein in Cell Extracts

Cells	ng MN/mg total protein
HeLa + MX	939.00
Rat2-Tk$^-$	1065.00
HeLa	27.50
XC	16.40
T24	1.18
HEF	0.00

The data were calculated from the results shown in Figure 12.

Example 9

Immunoelectron and Scanning Microscopy of Control and of MX-infected HeLa Cells

As indicated above in Example 1, MN antigen, detected by indirect immunofluorescence with MAb M75, is located on the surface membranes and in the nuclei of MX-infected HeLa cells or in HeLa cells grown in dense cultures. To elucidate more clearly the location of the MN antigen, immunoelectron microscopy was used wherein MAb M75 bound to MN antigen was visualized with immunogold beads. [Herzog et al., "Colloidal gold labeling for determining cell surface area," IN: Colloidal Gold, Vol. 3 (Hayat, M.A., ed.), pp. 139-149 (Academic Press Inc.; San Diego, CA).]

Ultrathin sections of control and of MX-infected HeLa cells are shown in Figure 13 A-D. Those immuno-electron micrographs demonstrate the location of MN antigen in the cells, and in addition, the striking ultrastructural differences between control and MX-infected HeLa. A control HeLa cell (Figure 13A) is shown to have on its surface very little MN antigen, as visualised with gold beads. The cell surface is rather smooth, with only two little protrusions. No mitochondria can be seen in the cytoplasm. In contrast, MX-infected HeLa cells (Figure 13B and C) show the formation of abundant, dense filamentous protrusions from their surfaces. Most of the MN antigen is located on those

filaments, which are decorated with immunogold. The cytoplasm of MX-infected HeLa contains numerous mitochondria (Figure 13C). Figure 13D demonstrates the location of MN antigen in the nucleus: some of the MN antigen is in nucleoplasm (possibly linked to chromatin), but a higher concentration of the MN antigen is in the nucleoli. Again, the surface of normal HeLa (panels A and E of Figure 13) is rather smooth whereas MX-infected HeLa cells have on their surface, numerous filaments and "blebs". Some of the filaments appear to form bridges connecting them to adjacent cells.

It has been noted that in some instances of <u>in vitro</u> transformed cells compared to their normal parent cells that one of the differences is that the surface of normal cells was smooth whereas on the transformed cells were numerous hair-like protrusions [Darnell et al. "Molecular Cell Biology," (2nd edition) Sci. Am. Books; W.H. Freeman and Co., New York (1990)]. Under that criteria MX-infected HeLa cells, as seen in Figure 13F, has a supertransformed appearance.

Further in some tumors, amplification of mitochondria has been described [Bernhard, W., "Handbook of Molecular Cytology," pp. 687-715, Lima de Faria (ed.), North Holland Publishing Co.; Amsterdam-London (1972)]. Such amplification was noted for MX-infected HeLa cells which

stained very intensely with Janus' green, specific for mitochondria whereas control HeLa were only weakly stained.

It should be noted that electron microscopists were unable to find any structural characteristics specific for tumor cells.

Example 10

Antisense ODNs Inhibit MN Gene Expression

To determine whether both of the p54/58N proteins were encoded by one gene, the following experiments with antisense ODNs were performed. Previously sparse-growing HeLa cells were seeded to obtain an overcrowded culture and incubated for 130 hours either in the absence or in the presence of two gene-specific ODNs complementary to the 5' end of MN mRNA. HeLa cells were subcultured at 8 x 10^5 cells per ml of DMEM with 10% FCS. Simultaneously, ODNs were added to the media as follows: (A) 29-mer ODN1 (5' CGCCCAGTGGGTCATCTTCCCCAGAAGAG 3' [SEQ. ID. NO.: 3], in 4 μM final concentration, (B) 19-mer ODN2 (5' GGAATCCTCCTGCATCCGG 3' [SEQ. ID. NO.: 4] in 4 μM final concentration and (C) both ODN1 and ODN2 in 2 μM final concentration each. (D) Cells treated in the same way, but incubated without ODNs, served as a control. After 130 hours, extracts from the cells were prepared and analyzed by immunoblotting using ^{125}I-labeled MAb M75. Protein extracts from the cells were

analyzed by immunoblotting and RIA using MAb M75. Figure 3 provides the immunoblot results of those experiments.

It was found that cultivation of HeLa cells with the ODNs resulted in considerable inhibition of p54/58N synthesis. The 19-mer ODN2 (Figure 3B) in 4 μM final concentration was very effective; as determined by RIA, it caused 40% inhibition, whereas the 29-mer ODN1 (4 μM) (Figure 3A) and a combination of the two ODNs (Figure 3C), each in 2 μM final concentration, were less effective in RIA showing a 25-35% increase of the MN-related proteins. At the same time, the amount of different HeLa cell protein determined by RIA using specific MAb H460 was in all cell variants approximately the same. Most importantly was that on immunoblot it could be seen that specific inhibition by the ODNs affected both of the p54/58N proteins. Thus, we concluded that the MN gene we cloned coded for both p54/58N proteins in HeLa cells.

The results indicated that the MN twin proteins arise by translation of a single mRNA (consistent with the Northern blotting data). Thus, the twin proteins may represent either differences in post-translational modification (phosphorylation, protease processing, etc.), or the use of alternative translational initiation sites.

Example 11

Northern Blotting of MN mRNA in Tumorigenic and Non-Tumorigenic Cell Lines

Figure 4 shows the results of Northern blotting of MN mRNA in human cell lines. Total RNA was prepared from the following cell lines by the guanidinium thiocyanate-CsCl method: HeLa cells growing in a dense (A) and sparse (B) culture; CGL1 (H/F-N) hybrid cells (C); CGL3 (D) and CGL4 (E) segregants (both H/F-T); and human embryo fibroblasts (F). Fifteen μg of RNA were separated on a 1.2% formaldehyde gel and blotted onto a Hybond C Super membrane [Amersham]. MN cDNA NotI probe was labeled by random priming [Multiprime DNA labelling system; Amersham]. Hybridization was carried out in the presence of 50% formamide at 42°C, and the final wash was in 0.1% SSPE and 0.1% SDS at 65°C. An RNA ladder (0.24-9.5 kb) [BRL; Bethesda, MD (USA)] was used as a size standard. Membranes were exposed to films at -70°C, with intensifying screens.

Detected was a 1.5 kb MN-specific mRNA only in two tumorigenic segregant clones--CGL3 and CGL4 (H/F-T), but not in the non-tumorigenic hybrid clone CGL1 (H/F-N) or in normal human fibroblasts. Further, the 1.5 kb mRNA was found in the HeLa cells growing in dense (Figure 4A) but not in sparse (Figure 4B) culture.

Thus, the results of the Northern blotting were consistent with other examples in regard to MN-related proteins being associated with tumorigenicity.

Example 12

Southern Blotting of Genomic DNAs from Different Vertebrate Species to Detect MN Gene and Restriction Analysis of Genomic DNA of HeLa Cells

Figure 5 illustrates the detection of MN genes in the genomic DNAs of various vertebrates by Southern blotting. Chromosomal DNA digested by PstI was as follows: (A) chicken; (B) bat; (C) rat; (D) mouse; (E) feline; (F) pig; (G) sheep; (H) bovine; (I) monkey; and (J) human HeLa cells. Restriction fragments were separated on a 0.7% agarose gel and alkali blotted onto a Hybond N membrane [Amersham]. The MN cDNA probe labelling and hybridization procedures were the same as for the Northern blotting analyses shown in Figure 4 and described in Example 11. The Southern blot of Figure 5 made with PstI indicates that the MN gene is conserved in a single copy in all vertebrate genomes tested.

HeLa. Further, genomic DNA from HeLa cells was prepared as described by Ausubel et al., Short Protocols in Molecular Biology [Greene Publishing Associates and Wiley-Interscience; New York (1989)], digested with different restriction enzymes, resolved on an agarose gel and transferred to Hybond N+ membrane [Amersham]. The HeLa

genomic DNA was cleaved with the following restriction enzymes with the results shown in Figure 17 (wherein the numbers in parentheses after the enzymes indicate the respective lanes in Figure 17): EcoRI (1), EcoRV (2), HindIII (3), KpnI (4), NcoI (5), PstI (6), and PvuII (7), and then analyzed by Southern hybridization under stringent conditions using MN cDNA as a probe.

The prehybridization and hybridization using an MN cDNA probe labelled with ^{32}P-dCTP by random priming [Multi-prime DNA labelling system; Amersham] as well as wash steps were carried out according to Amersham's protocols at high stringency. A 1 kb DNA Ladder [from BRL; Bethesda, MD (USA)] was used as a size standard. Membranes were exposed to films at -70°C, with intensifying screens.

The Southern blotting analysis of HeLa chromosomal DNA showed that the gene coding for MN is present in the human genome in a single copy (Figure 17). The sizes and distribution of MN-positive restriction fragments obtained using the restriction enzymes KpnI, NcoI and HindIII indicate that the MN gene contains introns, since those enzymes cut the MN genomic sequences despite the absence of their restriction sites in MN cDNA.

Example 13

Immunohistochemical Staining of Tissue Specimens

To study and evaluate the tissue distribution range and expression of MN proteins, the monoclonal antibody M75 was used to stain immunohistochemically a variety of human tissue specimens. The primary antibody used in these immunohistochemical staining experiments was the M75 monoclonal antibody. A biotinylated second antibody and streptavidin-peroxidase were used to detect the M75 reactivity in sections of formalin-fixed, paraffin-embedded tissue samples. A commercially available amplification kit, specifically the DAKO LSAB™ kit [DAKO Corp., Carpinteria, CA (USA)] which provides matched, ready made blocking reagent, secondary antibody and steptavidin-horseradish peroxidase was used in these experiments.

M75 immunoreactivity was tested according to the methods of this invention in multiple-tissue sections of breast, colon, cervical, lung and normal tissues. Such multiple-tissue sections were cut from paraffin blocks of tissues called "sausages" that were purchased from the City of Hope [Duarte, CA (USA)]. Combined in such a multiple-tissue section were normal, benign and malignant specimens of a given tissue; for example, about a score of tissue samples of breast cancers from different patients, a similar number of benign breast tissue samples, and normal breast tissue samples would be combined in one such multiple-

breast-tissue section. The normal multiple-tissue sections contained only normal tissues from various organs, for example, liver, spleen, lung, kidney, adrenal gland, brain, prostate, pancreas, thyroid, ovary, and testis.

Also screened for MN gene expression were multiple individual specimens from cervical cancers, bladder cancers, renal cell cancers, and head and neck cancers. Such specimens were obtained from U.C. Davis Medical Center in Sacramento, CA and from Dr. Shu Y. Liao [Department of Pathology; St. Joseph Hospital; Orange, CA (USA)].

Controls used in these experiments were the cell lines CGL3 (H/F-T hybrid cells) and CGL1 (H/F-N hybrid cells) which are known to stain respectively, positively and negatively with the M75 monoclonal antibody. The M75 monoclonal antibody was diluted to a 1:5000 dilution wherein the diluent was either PBS [0.05 M phosphate buffered saline (0.15 M NaCl), pH 7.2-7.4] or PBS containing 1% protease-free BSA as a protein stabilizer.

Immunohistochemical Staining Protocol

The immunohistochemical staining protocol was followed according to the manufacturer's instructions for the DAKO LSAB™ kit. In brief, the sections were dewaxed, rehydrated and blocked to remove non-specific reactivity as well as endogenous peroxidase activity. Each section was then incubated with dilutions of the M75 monoclonal

antibody. After the unbound M75 was removed by rinsing the section, the section was sequentially reacted with a biotinylated antimouse IgG antibody and streptavidin conjugated to horseradish peroxidase; a rinsing step was included between those two reactions and after the second reaction. Following the last rinse, the antibody-enzyme complexes were detected by reaction with an insoluble chromogen (diaminobenzidine) and hydrogen peroxide. A positive result was indicated by the formation of an insoluble reddish-brown precipitate at the site of the primary antibody reaction. The sections were then rinsed, counterstained with hematoxylin, dehydrated and cover slipped. Then the sections were examined using standard light microscopy. The following is an outline of exemplary steps of the immunohistochemical staining protocol.

1.	Series of ETOH-baths 100, 100, 95, 95, 70%	2 min. ± 1 min. each
2.	dH_20 wash - 2x	2 min. ± 1 min. each
3.	3% H_2O_2 as endogenous peroxidase block	5 min.
4.	PBS wash - 2x	2 min. ± 1 min.
5.	normal serum block (1.5% NGS)	30 min.
6.	primary antibody (Mab M75)	60 min. ± 5 min.
7.	PBS wash - 2x	2 min. ± 1 min.
8.	biotinylated secondary antibody	20-30 min. ± 2 min.
9.	PBS wash - 2x	2 min. ± 1 min.

10.	streptavidin-peroxidase reagent	20-30 min. ± 2 min.
11.	PBS wash - 2x	2 min. ± 1 min.
12.	DAB (150ml Tris, 90μl H_2O_2, 3ml KPL DAB)	5-6 min.
13.	PBS rinse, dH_2O wash	1-2 min.
14.	Hematoxylin counterstain	2 min. ± 1 min.
15.	wash with running tap water until clear	
16.	0.05% ammonium hydroxide	20 sec. ± 10 sec.
17.	dH_2O wash - 2x	3 min. ± 1 min.
18.	dehydrate 70, 95, 95, 100, 100% EtOH	2 min. ± 1 min. each
19.	xylene 3x	3 min. ± 1 min. each
20.	coverslip with Permount™ [Fisher Scientific Pittsburgh, PA (USA)]	
21.	wait 10 min. before viewing results.	

Interpretation. A deposit of a reddish brown precipitate over the plasma membrane was taken as evidence that the M75 antibody had bound to a MN antigen in the tissue. The known positive control (CGL3) had to be stained to validate the assay. Section thickness was taken into consideration to compare staining intensities, as thicker sections produce greater staining intensity independently of other assay parameters.

The above-described protocol was optimized for formalin-fixed tissues, but can be used to stain tissues prepared with other fixatives.

Results

Preliminary examination of cervical specimens showed that 62 of 68 squamous cell carcinoma specimens (91.2%) stained positively with M75. Additionally, 2 of 6 adenocarcinomas and 2 of 2 adenosquamous cancers of the cervix also stained positively. In early studies, 55.6% (10 of 18) of cervical dysplasias stained positively. A total of 9 specimens including both cervical dysplasias and tumors, exhibited some MN expression in normal appearing areas of the endocervical glandular epithelium, usually at the basal layer. In some specimens, whereas morphologically normal-looking areas showed expression of MN antigen, areas exhibiting dysplasia and/or malignancy did not show MN expression.

M75 positive immunoreactivity was most often localized to the plasma membrane of cells, with the most apparent stain being present at the junctions between adjacent cells. Cytoplasmic staining was also evident in some cells; however, plasma membrane staining was most often used as the main criterion of positivity.

M75 positive cells tended to be near areas showing keratin differentiation in cervical specimens. In some

specimens, positive staining cells were located in the center of nests of non-staining cells. Often, there was very little, if any, obvious morphological difference between staining cells and non-staining cells. In some specimens, the positive staining cells were associated with adjacent areas of necrosis.

In most of the squamous cell carcinomas of the cervix, the M75 immunoreactivity was focal in distribution, i.e., only certain areas of the specimen stained. Although the distribution of positive reactivity within a given specimen was rather sporadic, the intensity of the reactivity was usually very strong. In most of the adenocarcinomas of the cervix, the staining pattern was more homogeneous, with the majority of the specimen staining positively.

Among the normal tissue samples, intense, positive and specific M75 immunoreactivity was observed only in normal stomach tissues, with diminishing reactivity in the small intestine, appendix and colon. No other normal tissue stained extensively positively for M75. Occasionally, however, foci of intensely staining cells were observed in normal intestine samples (usually at the base of the crypts) or were sometimes seen in morphologically normal appearing areas of the epithelium of cervical specimens exhibiting dysplasia and/or malignancy. In such, normal appearing areas of cervical specimens, positive staining was seen in

focal areas of the basal layer of the ectocervical epithelium or in the basal layer of endocervical glandular epithelium. In one normal specimen of human skin, cytoplasmic MN staining was observed in the basal layer. The basal layers of these epithelia are usually areas of proliferation, suggesting the MN expression may be involved in cellular growth. In a few cervical biopsied specimens, MN positivity was observed in the morphologically normal appearing stratified squamous epithelium, sometimes associated with cells undergoing koilocytic changes.

Some colon adenomas (4 of 11) and adenocarcinomas (9 of 15) were positively stained. One normal colon specimen was positive at the base of the crypts. Of 15 colon cancer specimens, 4 adenocarcinomas and 5 metastatic lesions were MN positive. Fewer malignant breast cancers (3 of 25) and ovarian cancer specimens (3 of 15) were positively stained. Of 4 head and neck cancers, 3 stained very intensely with M75.

Although normal stomach tissue was routinely positive, 4 adenocarcinomas of the stomach were MN negative. Of 3 bladder cancer specimens (1 adenocarcinoma, 1 non-papillary transitional cell carcinoma, and 1 squamous cell carcinoma), only the squamous cell carcinoma was MN positive. Approximately 40% (12 of 30) of lung cancer specimens were positive; 2 of 4 undifferentiated carcinomas; 3 of 8 adenocarcinomas; 2 of 8 oat cell carcinomas; and, 5

of 10 squamous cell carcinomas. One hundred percent (4 of 4) of the renal cell carcinomas were MN positive.

In summary, MN antigen, as detected by M75 and immunohistochemistry in the experiments described above, was shown to be prevalent in tumor cells, most notably in tissues of cervical cancers. MN antigen was also found in some cells of normal tissues, and sometimes in morphologically normal appearing areas of specimens exhibiting dysplasia and/or malignancy. However, MN is not usually extensively expressed in most normal tissues, except for stomach tissues where it is extensively expressed and in the tissues of the lower gastrointestinal tract where it is less extensively expressed. MN expression is most often localized to the cellular plasma membrane of tumor cells and may play a role in intercellular communication or cell adhesion. Representative results of experiments performed as described above are tabulated in Table 3.

TABLE 3

Immunoreactivity of M75 in Various Tissues

TISSUE	TYPE	POS/NEG (#pos/#tested)
liver, spleen, lung, kidney, adrenal gland, brain, prostate, pancreas, thyroid, ovary, testis	normal	NEG (all)
skin	normal	POS (in basal layer) (1/1)
stomach	normal	POS
small intestine	normal	POS
colon	normal	POS
breast	normal	NEG (0/10)
cervix	normal	NEG (0/2)
breast	benign	NEG (0/17)
colon	benign	POS (4/11)
cervix	benign	POS (10/18)
breast	malignant	POS (3/25)
colon	malignant	POS (9/15)
ovarian	malignant	POS (3/15)
lung	malignant	POS (12/30)
bladder	malignant	POS (1/3)
head & neck	malignant	POS (3/4)
kidney	malignant	POS (4/4)
stomach	malignant	NEG (0/4)
cervix	malignant	POS (62/68)

The results recorded in this example indicate that the presence of MN proteins in a tissue sample from a patient may, in general, depending upon the tissue involved, be a marker signaling that a pre-neoplastic or neoplastic process is occurring. Thus, one may conclude from these results that diagnostic/prognostic methods that detect MN antigen may be particularly useful for screening patient samples for a number of cancers which can thereby be

detected at a pre-neoplastic stage or at an early stage prior to obvious morphologic changes associated with dysplasia and/or malignancy being evident or being evident on a widespread basis.

Example 14

Vaccine -- Rat Model

As shown above in Example 7, in some rat tumors, for example, the XC tumor cell line (cells from a rat rhabdomyosarcoma), a rat MN protein, related to human MN, is expressed. Thus a model was afforded to study antitumor immunity induced by experimental MN-based vaccines. The following representative experiments were performed.

Nine- to eleven-day-old Wistar rats from several families were randomized, injected intraperitoneally with 0.1 ml of either control rat sera (the C group) or with rat serum against the MN fusion protein GEX-3X-MN (the IM group). Simultaneously both groups were injected subcutaneously with 10^6 XC tumor cells.

Four weeks later, the rats were sacrificed, and their tumors weighed. The results are shown in Figure 14. Each point on the graph represents a tumor from one rat. The difference between the two groups -- C and IM -- was significant by Mann-Whitney rank test ($U = 84$, $\alpha < 0.025$). The results indicate that the IM group of baby rats developed tumors about one-half the size of the controls,

and 5 of the 18 passively immunized rats developed no tumor at all, compared to 1 of 18 controls.

Example 15

Expression of Full-Length MN cDNA in NIH 3T3 Cells

The role of MN in the regulation of cell proliferation was studied by expressing the full-length cDNA in NIH 3T3 cells. That cell line was chosen since it had been used successfully to demonstrate the phenotypic effect of a number of proto-oncogenes [Weinberg, R.A., Cancer Res., 49: 3713 (1989); Hunter, T., Cell, 64: 249 (1991)]. Also, NIH 3T3 cells express no endogenous MN-related protein that is detectable by Mab M75.

The full length MN cDNA was obtained by ligation of the two cDNA clones using the unique BamHI site and subcloned from pBluescript into KpnI-SacI sites of the expression vector pSG5C. pSG5C was kindly provided by Dr. Richard Kettman [Department of Molecular Biology, Faculty of Agricultural Sciences, B-5030 Gembloux, Belgium]. pSG5C was derived from pSG5 [Stratagene] by inserting a polylinker consisting of a sequence having several neighboring sites for the following restriction enzymes: EcoRI, XhoI, KpnI, BamHI, SacI, 3 times TAG stop codon and BglII.

The recombinant pSG5C-MN plasmid was co-transfected in a 10:1 ratio (10 μg : 1 μg) with the pSV2neo plasmid [Southern and Berg, J. Mol. Appl. Genet., 1: 327

(1982)] which contains the neo gene as a selection marker. The co-transfection was carried out by calcium phosphate precipitation method [Mammalian Transfection Kit; Stratagene] into NIH 3T3 cells plated a day before at a density of 1 x 10^5 per 60 mm dish. As a control, pSV2neo was co-transfected with empty pSG5C.

Transfected cells were cultured in DMEM medium supplemented with 10% FCS and 600 μg ml^{-1} of G418 [Gibco BRL] for 14 days. The G418-resistant cells were clonally selected, expanded and analysed for expression of the transfected cDNA by Western blotting using iodinated Mab M75.

For an estimation of cell proliferation, the clonal cell lines were plated in triplicates (2 x 10^4 cells/well) in 24-well plates and cultivated in DMEM with 10% FCS and 1% FCS, respectively. The medium was changed each day, and the cell number was counted using a hemacytometer.

To determine the DNA synthesis, the cells were plated in triplicate in 96-well plate at a density of 10^4/well in DMEM with 10% FCS and allowed to attach overnight. Then the cells were labeled with ^{3}H-thymidine for 24 hours, and the incorporated radioactivity was counted.

For the anchorage-independent growth assay, cells (2 x 10^4) were suspended in a 0.3% agar in DMEM containing

10% FCS and overlaid onto 0.5% agar medium in 60 mm dish. Colonies grown in soft agar were counted two weeks after plating.

Several clonal cell lines constitutively expressing both 54 and 58 kd forms of MN protein in levels comparable to those found in LCMV-infected HeLa cells were obtained. Selected MN-positive clones and negative control cells (mock-transfected with an empty pSG5C plasmid) were subjected to further analyses directed to the characterization of their phenotype and growth behavior.

The MN-expressing NIH 3T3 cells displayed spindle-shaped morphology, and increased refractility; they were less adherent to the solid support and smaller in size. The control (mock transfected cells) had a flat morphology, similar to parental NIH 3T3 cells. In contrast to the control cells that were aligned and formed a monolayer with an ordered pattern, the cells expressing MN lost the capacity for growth arrest and grew chaotically on top of one another (Figure 22a-d). Correspondingly, the MN-expressing cells were able to reach significantly higher (more than 2x) saturation densities (Table 4) and were less dependent on growth factors than the control cells (Figure 22g-h).

MN transfectants also showed faster doubling times (by 15%) and enhanced DNA synthesis (by 10%), as determined by the amount of [^{3}H]-thymidine incorporated in comparison

to control cells. Finally, NIH 3T3 cells expressing MN protein grew in soft agar. The diameter of colonies grown for 14 days ranged from 0.1 to 0.5 mm (Figure 22f); however, the cloning efficiency of MN transfectants was rather low (2.4%). Although that parameter of NIH 3T3 cells seems to be less affected by MN than by conventional oncogenes, all other data are consistent with the idea that MN plays a role in cell growth control.

Table 4

Growth Properties of NIH 3T3 Cells Expressing MN Protein

Transfected DNA	pSG5C/ pSV2neo	pSG5C-MN/ pSV2neo
Doubling time[a] (hours)	27.9 ± 0.5	24.1 ± 1.3
Saturation density[b] (cells x $10^4/cm^2$)	4.9 ± 0.2	11.4 ± 0.4
Cloning efficiency (%)[c]	< 0.01	2.4 ± 0.2

[a]For calculation of the doubling time, the proliferation rate of exponentially growing cells was used. [b]The saturation cell density was derived from the cell number 4 days after reaching confluency. [c]Colonies greater than 0.1 mm in diameter were scored at day 14. Cloning efficiency

was estimated as a percentage of colonies per number of cells plated, with correction for cell viability.

Example 16

Acceleration of G1 Transit and Decrease in Mitomycin C Sensitivity Caused by MN Protein

For the experiments described in this example, the stable MN transfectants of NIH 3T3 cells generated as described in Example 15 were used. Four selected MN-positive clones and four control mock-transfected clones were either used individually or in pools.

Flow cytometric analyses of asynchronous cell populations. For the results shown in Figure 23(a), cells that had been grown in dense culture were plated at 1 x 10^6 cells per 60 mm dish. Four days later, the cells were collected by trypsinization, washed, resuspended in PBS, fixed by dropwise addition of 70% ethanol and stained by propidium iodine solution containing RNase. Analysis was performed by FACStar using DNA cell cycle analysis software [Becton Dickinson; Franklin Lakes, NJ (USA)].

For the analyses shown in Figure 23(b) and (c), exponentially growing cells were plated at 5 x 10^5 cells per 60 mm dish and analysed as above 2 days later. Forward light scatter was used for the analysis of relative cell sizes. The data were evaluated using Kolmogorov-Smirnov test [Young, J. Histochem. Cytochem., 25: 935 (1977)]. D

is the maximum difference between summation curves derived from histograms. D/s(n) is a value which indicates the similarity of the compared curves (it is close to zero when curves are similar).

The flow cytometric analyses revealed that clonal populations constitutively expressing MN protein showed a decreased percentage of cells in G1 phase and an increased percentage of cells in G2-M phases. Those differences were more striking in cell populations grown throughout three passages in high density cultures [Figure 23(a)], than in exponentially growing subconfluent cells [Figure 23(b)]. That observation supports the idea that MN protein has the capacity to perturb contact inhibition.

Also observed was a decrease in the size of MN expressing cells seen in both exponentially proliferating and high density cultures. It is possible that the MN-mediated acceleration of G1 transit is related to the above-noted shorter doubling time (by about 15%) of exponentially proliferating MN-expressing NIH 3T3 cells. Also, MN expressing cells displayed substantially higher saturation density and lower serum requirements than the control cells. Those facts suggest that MN-transfected cells had the capacity to continue to proliferate despite space limitations and diminished levels of serum growth factors, whereas the control cells were arrested in G1 phase.

Limiting conditions. The proliferation of MN-expressing and control cells was studied both in optimal and limiting conditions. Cells were plated at 2 x 10^4 per well of 24-well plate in DMEM with 10% FCS. The medium was changed at daily intervals until day 4 when confluency was reached, and the medium was no longer renewed. Viable cells were counted in a hemacytometer at appropriate times using trypan blue dye exclusion. The numbers of cells were plotted versus time wherein each plot point represents a mean value of triplicate determination.

The results showed that the proliferation of MN expressing and control cells was similar during the first phase when the medium was renewed daily, but that a big difference in the number of viable cells occurred after the medium was not renewed. More than half of the control cells were not able to withstand the unfavorable growth conditions. In contrast, the MN-expressing cells continued to proliferate even when exposed to increasing competition for nutrients and serum growth factors.

Those results were supported also by flow cytometric analysis of serum starved cells grown for two days in medium containing 1% FCS. While 83% of control cells accumulated in G0-G1 phase (S = 5%, G2-M = 12%), expression of MN protein partially reversed the delay in G1 as indicated by cell cycle distribution of MN tranfectants (G0-G1 = 65%, S = 10%, G2-M = 26%). The results of the

above-described experiments suggest that MN protein might function to release the G1/S checkpoint and allow cells to proliferate under unfavorable conditions.

MMC. To test that assumption, unfavorable conditions were simulated by treating cells with the DNA damaging drug mitomycin C (MMC) and then following their proliferation and viability. The mechanism of action of MMC is thought to result from its intracellular activation and subsequent DNA alkylation and crosslinking [Yier and Szybalski, Science, 145: 55 (1964)]. Normally, cells respond to DNA damage by arrest of their cell cycle progression to repair defects and prevent acquisition of genomic instability. Large damage is accompanied by marked cytotoxicity. However, many studies [for example, Peters et al., Int. J. Cancer, 54: 450 (1993)] concern the emergence of drug resistant cells both in tumor cell populations and after the introduction of oncogenes into nontransformed cell lines.

The response of MN-transfected NIH 3T3 cells to increasing concentrations of MMC was determined by continuous [^{3}H]-thymidine labeling. Cells were plated in 96-well microtiter plate concentration of 10^4 per well and incubated overnight in DMEM with 10% FCS to attach. Then the growth medium was replaced with 100 μl of medium containing increasing concentrations of MMC from 1 μl/ml to 32 μg/ml. All the drug concentrations were tested in three

replicate wells. After 5 hours of treatment, the MMC was removed, cells were washed with PBS and fresh growth medium without the drug was added. After overnight recovery, the fractions of cells that were actively participating in proliferation was determined by continuous 24-hr labeling with [^{3}H]-thymidine. The incorporation by the treated cells was compared to that of the control, untreated cells, and the proliferating fractions were considered as a percentage of the control's incorporation.

The viability of the treated cells was estimated three days later by a CellTiter 96 AQ Non-Radioactive Cell Proliferation Assay [Promega] which is based on the bioreduction of methotrexate (MTX) into a water soluble formazan that absorbs light at 490 nm. The percentage of surviving cells was derived from the values of absorbance obtained after substraction of background.

The control and MN-expressing NIH 3T3 cells showed remarkable differences in their responses to MMC. The sensitivity of the MN-transfected cells appeared considerably lower than the control's in both sections of the above-described experiments. The results suggested that the MN-transfected cells were able to override the negative growth signal mediated by MMC.

ATCC Deposits. The material listed below was deposited with the American Type Culture Collection (ATCC) at 12301 Parklawn Drive, Rockville, MD 20852 (USA). The

deposits were made under the provisions of the Budapest Treaty on the International Recognition of Deposited Microorganisms for the Purposes of Patent Procedure and Regulations thereunder (Budapest Treaty). Maintenance of a viable culture is assured for thirty years from the date of deposit. The hybridomas and plasmids will be made available by the ATCC under the terms of the Budapest Treaty, and subject to an agreement between the Applicants and the ATCC which assures unrestricted availability of the deposited hybridomas and plasmids to the public upon the granting of patent from the instant application. Availability of the deposited strain is not to be construed as a license to practice the invention in contravention of the rights granted under the authority of any Government in accordance with its patent laws.

Hybridoma	Deposit Date	ATCC #
VU-M75	September 17, 1992	HB 11128
MN 12.2.2	June 9, 1994	HB 11647

Plasmid	Deposit Date	ATCC #
A4a		
XE1		
XE3		

The description of the foregoing embodiments of the invention have been presented for purposes of

illustration and description. They are not intended to be exhaustive or to limit the invention to the precise form disclosed, and obviously many modifications and variations are possible in light of the above teachings. The embodiments were chosen and described in order to explain the principles of the invention and its practical application to enable thereby others skilled in the art to utilize the invention in various embodiments and with various modifications as are suited to the particular use contemplated.

All references cited herein are hereby incorporated by reference.